300 μL of H2O was then added to the tube, and the cells were fully lysed by an ultrasonic cell disruptor (Bioruptor Pico, Diagenode, Belgium) for 10 min at 4°C. After mixing, 20 μL of suspension was collected for protein quantification and subsequent data normalization. Then, methanol (precooled to −80°C) was added into the tube to make 80% methanol solution (v/v). The tube was vortexed vigorously for 5 min and centrifuged at 14,000 × g for 10 min at 4°C to remove the cell debris. Afterward, the metabolite-containing supernatant was transferred to a new tube on ice. The supernatant was evaporated to dryness in a vacuum concentrator and reconstituted in 100 μL of methanol: H2O (1:1, v/v), which was centrifuged again at 14000 × g for 15 min at 4°C to remove insoluble debris.